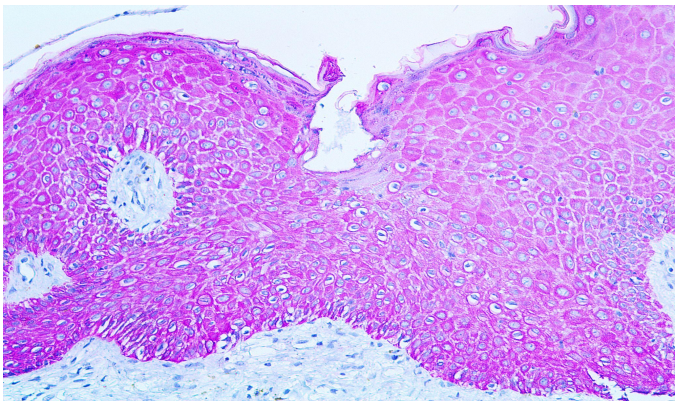
PolyDetector HRP Fuchsia Substrate-Chromogen

IVD



Inset: IHC of AE1/AE3 on a FFPE skin tissue using HRP Fuchsia Substrate-Chromogen.

Intended Use
For In Vitro Diagnostic Use.

Summary and Explanation
PolyDetector HRP Fuchsia Substrate-Chromogen is suitable for use with peroxidase detection systems and allows for the demonstration of cell antigens or nucleic acids in paraffin-embedded tissues, cryostat sections, cytosmears, and cell preparations. The substrate-chromogen is the final step in the detection portion; it enables the antibody antigen complex to be viewed under the light microscope. Ideal for multiplex IHC or CISH purposes. This substrate-chromogen is stable after mixing for up to 24 hours, which makes it ideal for use in automated stainers, and particularly useful in tissues where there is endogenous melanin.

HRP Fuchsia forms a permanent record of the stain results when coverslipped with PermaMounter (BSB 0094-BSB 0097) or XyGreen PermaMounter (BSB 0169 – BSB 0174). It can also be mounted with media such as AquaMounter (BSB 0090-BSB 0093).

Presentation
HRP Fuchsia is a chromogen (color forming molecule) that develops into a vibrant fuchsia/pink color.

Catalog No.	*Buffer-Substrate*	*Chromogen*
BSB-0364-15	15 ml	1 ml
BSB-0364-50	50 ml	3 ml
BSB-0364-100	100 ml	6 ml
BSB-0364-200	200 ml	12 ml
BSB-0364-500	500 ml	25 ml
BSB-0364-1000	1000 ml	50 ml

Storage Store at 2-8°C

Stability
This product is stable up to the expiration date on the product label.
Do not use after the expiration date listed on package label. Temperature fluctuations should be avoided. Store appropriately when not in use, and avoid prolonged exposure to room temperature conditions.

Abbreviated Immunohistochemical Protocol

Step	ImmunoDetector HRP	PolyDetector HRP	PolyDetector Plus HRP
Peroxidase Blocker	5 min	5 min	5 min
Primary Antibody	30-60 min.	30-60 min.	30-60 min.
1st Step Detection	10 min.	30-45 min.	15 min.
2nd Step Detection	10 min.	N/A	15 min.
Substrate-Chromogen	5-10 min.	5-10 min.	5-10 min.
Counterstain / Coverslip	Varies	Varies	Varies

Preparation of a Working Solution.
To prepare a working PolyDetector HRP Fuchsia Substrate-Chromogen solution, add 2 drops of Chromogen to 1 mL of the HRP Fuchsia Buffer-Substrate. Mix the two solutions well.

1 drop of HRP Fuchsia chromogen equals ~ 25 ul.

Working HRP Fuchsia Substrate Chromogen Required	1 ml	2 ml	3 ml
HRP Fuchsia Buffer	1 ml	2 ml	3 ml
HRP Fuchsia Chromogen	2 drops	4 drops	6 drops

Counterstaining
HRP Fuchsia can be counterstained for 30-60 seconds with Hematoxylin (BSB 0024 - BSB 0028). For detailed counterstaining protocols refer to PI 0028.

Mounting Protocol
Aqueous Mounting Protocol
1. After the histological, immunohistochemical or in situ hybridization staining procedure is completed, rinse slides with a TBST buffer (BSB 0042).
2. Apply 1-3 drops of an Aqueous Mounting medium such as AquaMounter (BSB 0090-0093) or similar mounting media.
3. Apply cover slip and air dry at room temperature before microscopic observation.
Perfect for immediate observation.

Permanent Mounting Protocol

a. Alcohol/Xylene Protocol

1. After the histological, immunohistochemical or in situ hybridization staining procedure is completed, rinse slides in deionized water.
2. Dip the slides in alcohol 30%, 70%, and 100% for 1-2 minutes, then dip for 1-2 minutes in 3 xylenes.
3. Add an organic Permanent Mounting medium such as XyGreen PermaMounter (BSB 0169-0174), PermaMounter (BSB 0094-0097) or similar permanent mounting media.
4. Apply cover slip and air dry before microscopic observation.

b. ChromoProtector Protocol

1. After the histological, immunohistochemical or in situ hybridization staining procedure is completed, rinse slides in deionized water. Do not incubate tissue or cell
specimens in solvents such as alcohol, toluene, or xylene.
2. Using a coplin jar, or a staining dish with a rack, immerse slides with tissues in ChromoProtector or lay wet slides horizontally and apply sufficient drops of
ChromoProtector (BSB 0151 – BSB 0156) to completely cover the tissue. Carefully spread ChromoProtector if needed, but avoid contacting the tissue.
3. Incubate slides for ten minutes at 60 °C to allow ChromoProtector to penetrate tissues.
4. Remove excess ChromoProtector by placing slides vertically over an absorbent material and let excess drain off into absorbent material. Do not rinse slides.
5. Allow slides to COMPLETELY air dry.
NOTE: The ChromoProtector will protect tissue from drying artifacts during the air-drying process.
6. When slides are completely dried, they can be mounted using most standard mounting methods such as aqueous or permanent.

c. Fast ChromoProtector Protocol

- Fast ChromoProtector with Cap Gap

If using the Bio SB Capillary Gap system, draw the Fast ChromoProtector up into the capillary gap, completely covering the tissue. Allow it to sit for 30 seconds to 1 minute, then separate the slides and allow the Fast ChromoProtector to drain off. Dry the slides completely and mount with one drop of mounting media.

- Fast ChromoProtector on single slides

Drip the Fast ChromoProtector on the tissue, allow it to sit for 30 sec. and let it drain off the tilted slide. Dry the slides completely and mount with one drop of mounting media.
After Fast ChromoProtector pretreatment, mount a slide with aqueous or permanent mounting media.

Product Limitations

Due to inherent variability present in immunohistochemical procedures (including fixation time of tissues, dilution factor of antibody, retrieval method utilized, and incubation time), optimal performance should be established through the use of positive and negative controls. Results should be interpreted by a qualified medical professional.

Precautions

1. For professional users only. Results should be interpreted by a qualified medical professional.
Ensure proper handling procedures are used with this reagent.
2. Always wear personal protective equipment such as a laboratory coat, goggles, and gloves when handling reagents.
3. Dispose of unused solution with copious amounts of water.
4. Do not ingest reagents. If the reagent is ingested, seek medical advice immediately.
5. Avoid contact with eyes. If contact occurs, flush with large quantities of water.
6. Follow safety precautions of the heating device used for epitope retrieval (TintoRetriever Pressure Cooker or similar).
7. For additional safety information refer to the Safety Data Sheet for this product.
8. For complete recommendations for handling biological specimens, please refer to the CDC document, "Guidelines for Safe Work Practices in Human and Animal Medical Diagnostic Laboratories" (see References in this document).

References

1. U.S. Department of Health and Human Services: Centers for Disease Control and Prevention. Guidelines for Safe Work Practices in Human and Animal Medical Diagnostic Laboratories. Supplement / Vol. 61, January 6, 2012.

Symbol Key/Légende des symboles/Erläuterung der Symbole

	Storage Temperature Limites de température Zulässiger Temperaturbereich	Manufacturer Fabricant Hersteller	REF Catalog Number Référence du catalogue Bestellnummer
IVD In Vitro Diagnostic Medical Device Dispositif médical de diagnostic in vitro In-Vitro-Diagnostikum	Read Instructions for Use Consulter les instructions d'utilisation Gebrauchsanweisung beachten	Expiration Date Utiliser jusque Verwendbar bis	LOT Lot Number Code du lot Chargenbezeichnung

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