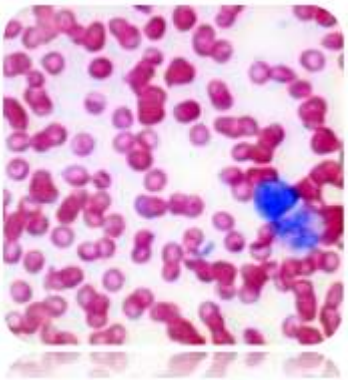
SS1044-VO

Wright-Giemsa Stain Kit

Description: Wright-Giemsa Stain Kit is intended to be used for differential staining of blood smears, bone marrow and blood parasites.

Erythrocytes:	Pink-Tan
Leukocytes:	Blue-Purple
Neutrophils:	Light Purple or Lavender granules in cytoplasm.
Eosinophils:	Bright Red or Red-Orange granules in cytoplasm.
Basophils:	Deep Purple or Violet-Black granules in cytoplasm.
Platelets:	Violet-Purple granules in light blue cytoplasm.

Uses/Limitations:
For In-Vitro Diagnostic use only.
Hematology applications.
Do not use past expiration date.
Use caution when handling these reagents.



Control Tissue: Blood smear on clean slide.

Kit Contents:

Description	Volume	StorageConditions
Wright-Giemsa Solution	500 ml	Room Temperature
Phosphate Buffer Solution (pH 6.8)	500 ml (x2)	Room Temperature

Required but not included:

Description	Volume	StorageConditions
Methanol, Absolute	500 ml	Room Temperature
Water, Deionized/Distilled	1 Gallon	Room Temperature

Precautions:
Avoid contact with skin and eyes.
Flammable.
May be fatal or cause blindness if swallowed.
Poison.
Follow all Federal, State, and local regulations regarding disposal.
Use in chemical fume hood whenever possible.

Storage: 18° C 25° C

Revision Date: 03/29/2016

Cancer Diagnostics, Inc. 116 Page Point Circle
Durham, NC 27703 USA
(877) 846 5393 www.cancerdiagnostics.com

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Preparation of Reagents Prior to Beginning:

1. Prepare **Working Wright-Giemsa Solution** by mixing 25ml of Wright-Giemsa Solution with 25ml of Phosphate Buffer Solution, pH 6.8 .

Procedure (Standard):

1. Smear a small drop of blood on a clean microscope slide and allow to air dry.
2. Fix by placing in absolute Methanol for 5 minutes.
3. Place slide in staining tray and flood with Working Wright-Giemsa Solution for 5 minutes. Note: Agitate slide occasionally to insure proper staining.
4. Rinse slide in deionized/distilled water.
5. Flood slide with Phosphate Buffer Solution, pH 6.8 until no stain runs off.
6. Allow slide to remain in Phosphate Buffer Solution, pH 6.8 for an additional 1 minute.
7. Dip slide in distilled water and air dry at room temperature.
8. Dip slide several times in Xylene or Xylene Substitute.
9. Mount in synthetic resin.

References:

1. Sheehan, D., Hrapchak, B., Theory and Practice of Histotechnology: 2nd Edition, 1980, pages 155-156.

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