SS1033-250

May-Grunwald Giemsa Stain Kit

Solutions Provided:
SSC1207-250 - **Jenner Stain Stock Solution**
SSC1149-250 - **Giemsa Stain Stock Solution**
SSC1208-250 - **Giemsa Buffer, pH 6.8**
SSC1031-250 **- Acetic Acid, 1%**

Working solution preparation (*prepare just before use*):

Jenner Stain Working Solution		**Giemsa Stain Working Solution**	
Jenner Stain Stock Solution:	25mL	Giemsa Stain Stock Solution:	2.5mL
Giemsa Buffer, pH 6.8*:	25mL	Giemsa Buffer, pH 6.8*:	50mL

*Final pH of working stain solutions will produce varying results. User preference may require the use of distilled/deionized water or the Giemsa Buffer, pH 7.2.

Conventional Procedure

1. Deparaffinize and hydrate sections through alcohol to distilled water.
2. Place sections in 100% methanol for 3 minutes.
3. Place sections in a second change of 100% methanol for 3 minutes.
4. Stain sections in **Jenner Stain Working Solution** for **5-6 minutes**.
5. Stain sections in **Giemsa Stain Working Solution** for **45 minutes**.
6. Rinse slides briefly in distilled water.
7. Dip slides in **Acetic Acid, 1%** and check under microscope until desired differentiation has been achieved.
8. Rinse slides in distilled water
9. Dehydrate rapidly in 3 changes of 100% alcohol.
10. Clear and mount with appropriate mounting medium.

Results:

Nuclei:	Blue
Cytoplasm:	Shades of pink
Bacteria:	Blue

Notes:

1. After initial deparffination and hydration: If sections have been fixed in Zenker, B-5 Solution or any other solution containing mercury be sure to prepare slides by placing in Gram's or Lugol's Iodine for 10 minutes, washing in tap water followed by Sodium Thiosulfate, 5% for 5 minutes. After one final wash in tap water and rinse in distilled water, proceed to **Step 2**.

Revision Date: 12/29/2016

Cancer Diagnostics, Inc. 4300 Emperor Blvd - 400
Durham, NC 27703 USA
(877) 846 5393 www.cancerdiagnostics.com