



SS1026-MG-VO

Trichrome Stain Kit

(Modified Gomori's)

Description:

The Trichrome Stain Kit (Modified Gomori's) is intended for use in the histological visualization of collagenous connective tissue fibers in tissue sections. The Gomori Trichrome is a more simplified procedure than other more traditional trichromes. Additionally, this procedure is useful in the assessment of the degree of fibrosis in liver biopsies.

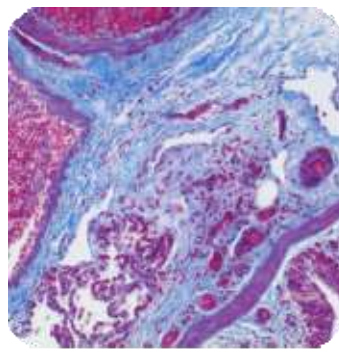
Collagen:	Blue
Muscle Fibers:	Red
Nuclei:	Black/Blue

Uses/Limitations:

Not to be taken internally.
For In-Vitro Diagnostic use only.
Histological applications.
Do not use if reagents become cloudy.
Do not use past expiration date.
Use caution when handling reagents.
Non-Sterile.

Control Tissue:

Lung
Uterus
Small Intestine
Appendix

**Availability/Contents:**

<u>Kit Contents</u>	<u>Volume</u>	<u>Storage</u>
Bouin's Fluid	125 ml	18-25°C
Hematoxylin, Weigert's Iron (Part A)	125 ml	18-25°C
Hematoxylin, Weigert's Iron (Part B)	125 ml	18-25°C
Trichrome Stain Solution (Blue)	125 ml	18-25°C
Acetic Acid Solution (0.5%)	250 ml	18-25°C
Acid Alcohol Solution (0.5%)	250 ml	18-25°C

Precautions:

Keep away from open flame.
Avoid contact with skin and eyes.
Harmful if swallowed.
Follow all Federal, State, and local regulations regarding disposal.
Use in chemical fume hood whenever possible.

Storage: 18° C  25° C





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Procedure:

1. Deparaffinize sections if necessary and hydrate to distilled water.
2. Preheat Bouin's Fluid in a water bath to 62° - 64° centigrade in a fume hood or very well ventilated area.
3. Place slides in preheated Bouin's Fluid for 60 minutes followed by a 10 minute cooling period.
4. Rinse slide in tap water until section is completely clear.
5. Rinse 1 time in distilled water.

Note: Use mixed Hematoxylin, Weigert's Iron immediately and dispose of directly thereafter.

6. Mix an equal volume of Hematoxylin, Weigert's Iron (Parts A and B) and apply to tissue section for 5 minutes.
7. Rinse slide in tap water for 2 minutes.
8. Apply Acid Alcohol Solution (0.5%) to slide for 5 seconds to adjust pH and remove excess stain.
9. Rinse slide in tap water for 1 minute.
10. Rinse slide 1 time in distilled water.
11. Apply Trichrome Stain (Blue) to tissue section and incubate for 15 minutes.
12. Rinse slide quickly in distilled water.
13. Apply Acetic Acid Solution (0.5%) to tissue section for 10 seconds.
14. Rinse slide using absolute alcohol.
15. Dehydrate in 2 changes of absolute alcohol, clear, and mount in synthetic resin.

References:

1. Gomori, G.L., A rapid one-step trichrome. Amer J of Clinical Pathology. Vol. 20: page 661, 1950.
2. Manual of Histologic Staining Methods of AFIP. 3rd Edition, McGraw-Hill Publishing. Page 93, 1968.
3. Elbadawi, A., Hexachrome modification of Movat's stain. Stain Technology, Vol. 51: pages 249-253, 1976.
4. Carson, F.L., Histotechnology; A Self-Instructional Text, 2nd Edition. ASCP Press, Chicago, IL. Pages 117-121, 1996.
5. Tretheway, D., et al. Should Trichrome Stain Be Used on All Post-Liver Transplant Biopsies with Hepatitis C Virus Infection To Estimate the Fibrosis Score? Liver Transplantation, May; 14(5): pages 695-700, 2008.

Storage: 18° C  25° C



**SS1026-MG-VO**

Description:	Volume
Bouin's Fluid	125 ml 500 ml 1000 ml
Hematoxylin, Weigert's Iron Kit (Two Components)	250 ml 1000 ml 2000 ml
Trichrome Stain Solution (Blue)	125 ml 500 ml 1000 ml
Acetic Acid Solution (0.5%)	250 ml 500 ml 1000 ml
Acid Alcohol Solution (0.5%)	250 ml 500 ml 1000 ml

Storage: 18° C  25° C

